

Vascular plant species inventory of a Philippine lowland rain forest and its conservation value

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Abstract. The Philippines are one of the most important biodiversity hotspots on earth. Due to the extraordinary rate of environmental destruction, leaving only 3% of the land with primary forest, this biodiversity is at high risk. Despite that situation information on Philippine forest vegetation is fragmentary and focused on trees. This study aimed at analysing forest remnants in the Leyte Cordillera on the Island of Leyte, and at evaluating their role as refuge to the largely destroyed lowland forest vegetation. A total of 49 plots (100 m² each) between 55 and 520 m a.s.l. were studied. All vascular plant species except epiphytes were included. Records include 685 taxa from 289 genera and 111 families, representing nearly 8% of the known Philippine vascular plant species. More than half (52%) of the species are Philippine endemics. A number of 41 tree species, or 6% of all taxa recorded, are included in the IUCN red list, either as vulnerable, endangered, or critically endangered. Life form composition was dominated by phanerophytes (65.3%), followed by lianas and chamaephytes (17.1 and 16.9%, respectively). The most common families were the Rubiaceae with 35 and the Euphorbiaceae with 32 species. All five Philippine dipterocarp forest types as well as the molave forest type were represented by typical tree species. The area provides an important gene bank of the highly threatened Philippine lowland forest vegetation and is of high value for biodiversity conservation. Additionally, it can play an important role as seed source of valuable tree species for the increasing initiatives to rehabilitate and reforest degraded land with native species.

Introduction

The destruction of tropical rain forests is still continuing at high rates (FAO 2003). This process, especially threatens the earth's biodiversity hotspots such as the Philippines (Myers et al. 2000; Brooks et al. 2002). Despite this, there are only very few studies worldwide which aimed at the documentation of the total plant species richness of such sites. Most inventories were restricted to selected life forms such as ground herbs (e.g. Kiew 1987; Poulsen and Balslev 1991; Poulsen 1996) or trees of a defined minimum diameter (e.g. Valencia et al. 1994; Lieberman et al. 1996; Newbery et al. 1996; Rennolls and Laumonier 2000; Slik et al. 2003). Vascular plant species composition of tropical lowland forests was studied in Ghana on 0.5- and 1-ha plots by Hall and Swaine (1981), in Amazonia on 0.02-ha plots by Takeuchi (1960) and on 10 non-contiguous plots of 0.1 ha by Duivenvoorden

(1994), in Ecuador on 0.1-ha plots of three lowland forest types by Gentry and Dodson (1987), and in stratified plots with a total area of about 2 ha in Puerto Rico by Smith (1970). In Southeast Asia, Kochummen et al. (1992) studied the trees and shrubs (> 1 cm diameter at breast height (dbh)) in a 50-ha plot in the Pasoh Forest Reserve in Malaysia. The most comprehensive study including all vascular plants as well as mosses was conducted by Whitmore et al. (1985) on a single 100-m² plot in the lowland rain forest of Costa Rica. However, no study representing a complete inventory of vascular plant species richness of any site of lowland rain forest in Southeast Asia was found.

The Philippines are among the most seriously depleted tropical countries with only 3% of the land area still covered by primary forest (Myers et al. 2000). From 1990 to 2000, the Philippines lost 1.4%, or 89,000 ha, of the forest area annually (FAO 2003). At the same time, the Philippine archipelago is one of the most important biodiversity hotspots on earth (Myers et al. 2000) with high proportions of endemic plant and animal species (Heaney and Regalado 1998). The endemism rate of plants was estimated to be 39% (Davis et al. 1995), but for certain taxa, it can be much higher. For example, 11 of 12 species of pitcher plants (*Nepenthes* spp.) known from the Philippines are endemic (Cheek and Jebb 2001). Similarly, there are high rates of endemism among the fauna. Referring to terrestrial vertebrates, 64% of the archipelago's land mammals are endemic, as well as 44% of the breeding land birds, 68% of the reptiles, and ca. 78% of the amphibians (Heaney and Regalado 1998). Most of them depend on forest ecosystems.

Despite the ecological uniqueness on the one hand and the extensive destruction on the other, the study of Philippine forest vegetation has been neglected (Tan and Rojo 1989; Kartawinata 1990; Soerianegara and Lemmens 1994). Much of the current knowledge is still based on studies conducted in the early 20th century (Whitford 1906; Whitford 1911; Brown and Mathews 1914; Brown 1919), which were mainly dealing with timber trees under economical aspects. Recent studies focused on the vegetation of montane and submontane forest types on different islands. However, in most cases (Aragones 1991; Pipoly and Madulid 1998; Proctor et al. 1998; Hamann et al. 1999) these were largely restricted to trees of a defined size, which usually is ≥ 10 cm dbh. Buot and Okitsu (1997) only considered woody plants higher than 1.3 m, and Ingle (2003) those of at least 5 cm dbh. The only data without size limitations are provided by Gonzales-Salcedo (2001) from Mt. Amuyao, Luzon, at elevations of 1600–1800 a.s.l. and by Gruezo (1998) from the highly degraded vegetation of Pagbilao Grande Island. No study dealing with lowland forest vegetation was found in the literature.

In order to provide more substantial information on species richness and composition of Philippine lowland forests, we analysed forest remnants in the rugged foothills of the Leyte Cordillera. The island of Leyte is located in the central part of the Philippine archipelago and represents a typical example of the environmental situation in the Philippines. In 1987, the remaining forest cover of Leyte was 12%, and in 1994 only 2% of the island's area have been

estimated to be primary forest (Dargantes and Koch 1994). More recent data (DENR 1998) show that about 40% of the land area of Leyte is covered by grassland and barren land, resulting from abandoned cultivation and grazing land that marginalised in productivity through erosion and leaching. Another 40% of the island's area is under coconut plantations. The remaining area is composed of settlements, agricultural land and forest. In the view of this situation, the objectives of this study were (a) to analyse the vascular plant species composition and diversity of selected plots of mature primary forest and (b) to evaluate the role of the study area as refuge to lowland forest vegetation and its significance for conservation and as a gene bank.

Material and methods

Study area

The Island of Leyte (Figure 1) belongs to the biogeographic region of the Eastern Visayas (DENR and UNEP 1997). It is located between 9°55' N–11°48' N and 124°17' E–125°18' E, with an extension of 214 km from north to south. Located offshore the northeastern part of Leyte is the island of Samar. The southern part of Leyte is exposed to the Pacific Ocean (Leyte Gulf).

Leyte is characterised by the north–south running Leyte Cordillera which is part of the Philippine Fault Line. The Cordillera reaches a maximum elevation of ca. 1350 m (Mt. Lobi) in the northern part of the island. As geologically young volcanic mountain range, it shows a typical rugged topography of narrow ridges and steep slopes, where landslides are common (Bremer 1995, 1999; Walsh 1996). In its foothills, patches of primary forest without discernible human interference can still be found although the island is densely populated (ca. 262 inhabitants/km² as calculated after: NSCB RU-8 2001). The coastal plains have already been deforested in the first half of the last century (Barrera et al. 1954).

The study area is located in the western part of the Leyte Cordillera, ca. 8 km north of the provincial capital of Baybay, in the foothills of Mt. Pangasugan (1150 m, 10°46' N, 124°50' E). In this part of the island, the Cordillera reaches close to the coast. Large parts of the mountain's western range are extremely steep and are free from trees of this reason. In the eastern part of the Cordillera, the slope has a lower gradient.

Primary forest can be found from about 250 m a.s.l. up to the mountain's summit at 1150 m a.s.l. In hillsides below this elevation, the forests has largely been replaced by coconut plantations and slash and burn agriculture. Only along the small creeks, near natural vegetation can still be found at these lower elevations. Above ca. 600 m a.s.l., the lowland forest formation changes into mossy forest, with its stunted trees and a rich epiphyte community.

Within the study area, no recent logging was observed, although forest clearing continues at other localities of the mountain. This can be explained by the area's status as Forest Reserve of the Leyte State University. Despite this, rattan collection and hunting was observed within the Forest Reserve.

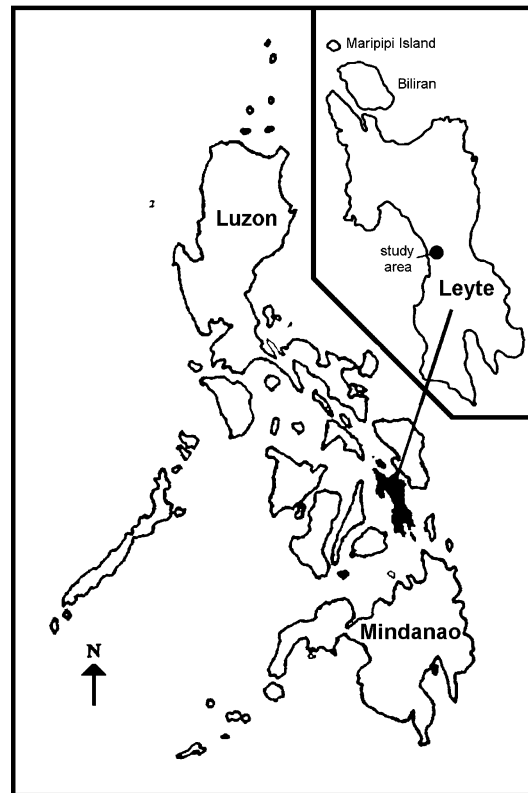


Figure 1. The Philippine archipelago, the Island of Leyte, and the location of the study area.

Geology and soils

The soil type in the primary forest between 370 and 520 m a.s.l. is a haplic Andosol with rudic phase (FAO/UNESCO 1988) overlying basaltic and andesitic breccia (Zikeli 1998). The soil at lower elevations (100 m a.s.l.) has been classified as haplic Alisol (FAO/UNESCO 1988) over basalt (Asio 1996).

Climate

According to the climatic classification of the 'Modified Corona's System', Leyte is climatically divided (Kintanar 1984). Southern Leyte belongs to the climatic type II (i.e. no pronounced dry season), but exhibits a distinct rainfall peak in December and January as a result of the northeast monsoon. The northern part of Leyte which includes the study area has been assigned to climatic type IV, showing a more or less even rainfall distribution throughout

the year. The standardization of the rainfall pattern in northern Leyte compared to that of southern Leyte might be explained by the protective effect of the Island of Samar off the northeast-coast of Leyte, although Samar's mountains are lower than those of Leyte (ca. 850 m a.s.l.).

Local climatic conditions have been analysed from a 23-years period (1976–1998) of record by using data from the PAGASA (Philippine Atmospheric, Geophysical and Astronomical Services Administration) weather station on the campus of the Leyte State University (7 m a.s.l.), ca. 1–3 km west of the study plots. The annual average temperature is 27.4 °C and the average annual precipitation is 2586 mm. Highest precipitation occurs during November to January. Lowest rainfall is observed between March and May. On average, all months receive at least 100 mm precipitation, i.e. there is no dry month according to the definition of Walsh (1996). However, drought periods (i.e. less than 50 mm of monthly precipitation according to Walsh 1996) of up to 4 months have been recorded during El Niño Southern Oscillation events. The general rainfall patterns and the climatic conditions measured at the PAGASA station are more similar to climatic type II with its clear impact of the northeast monsoon than to climatic type IV, implying that neither the mountains of Samar nor the Leyte Cordillera itself causes a distinct rain shadow west of Mt. Pangasugan.

Orographic rainfalls are an important factor in the Leyte Cordillera, especially in the vicinity of the mountain summits. The summit of Mt. Pangasugan is often observed being cloud covered, and during field work heavy rainfalls have been experienced, while the coastal plain did not receive any precipitation.

An important climatic feature of the area are typhoons. Leyte lies at the southern margin of the typhoon tracks entering the Philippines, and is hit at a rate of five typhoons in three years, mainly during the summer months (Parong 1984; cited in Kintanar 1984).

Vegetation analysis

Field studies were conducted in 1997 and 1998. The attempt to identify a minimum area in mature primary forest failed due to the heterogeneity of the vegetation. A plot size allowing a reasonable number of replications proved to be 100 m². Where relief conditions and homogeneity of the vegetation allowed, plots were arranged along a catena from ridge to river bank. The 100-m² plots were generally designed as quadrats, but on narrow ridges and river banks, other rectangular design was used due to relief constraints. A total of 49 plots was established, with 15 on the ridge, 21 on slopes, and 11 on riverbanks. Two plots were established in ca. 6-year-old land slide successions, with one of them located ca. 2 km south of the main study area.

The vegetation analysis procedure was based on a 'nested quadrat design' (Kent and Coker 1992). All plants >2.5 m were identified from the total plots (100 m²). On central subplots of 25 m², all plants ≤ 2.5 m

as well as the lianas were considered. Records included epiphytic and climbing plants on the stem bases of trees up to a height of 2.5 m. Crown epiphytes were not included in the analysis, but epiphytes found on the ground were identified and added to the species lists. From species which could not be identified in the field, voucher specimens were collected. Tall trees were sampled with the help of a tree climber. However, no samples were taken from erect and climbing palms (rattans), because this would have been destructive and the chance of identification was very low due to the lack of fertile specimens. Therefore, most palms had to be distinguished as morphospecies.

Taxa were assigned to life forms on the base of field observations, or with the help of literature information in the case of juveniles. Life form classification followed Ellenberg and Mueller-Dombois (1967). Plant samples collected in this study were deposited at the Department of Plant Breeding, Herbarium, Leyte State University, ViSCA, Baybay, Leyte, 6521A, Philippines.

Species identification and nomenclature

Identification of specimens was conducted with the help of literature and specialists. Publications referring to the Philippine flora included de Guzman et al. (1986), Merrill (1912), Pancho (1983), Santos et al. (1986), van Steenis (1950-ongoing), and Zamora and Co (1986). Sources referring to neighbouring countries, which also include many Philippine taxa, were used in addition (Henderson 1974a, b; Ng 1978, 1989; Keng 1983, 1990; Whitmore 1983a, b; Corner 1988; Anonymous 1993, 1994, 1996; Soepadmo and Wong 1995; Soepadmo et al. 1996). Identification of seedlings, infertile and juvenile plants was not possible in all cases. For pre-identification of taxa and delimitation of morphospecies, field characters were very important. Besides the description of such characters in the above mentioned literature, the specific publications by van Balgooy (1997a, b) and Keller (1996) were used. Tree seedlings were identified with help of Ng (1991) and Burger (1972).

In addition, plants were identified by taxonomists from the Philippine National Herbarium, Manila, the National Herbarium of the Netherlands, Leiden Branch, and collaboratively during meetings of the Philippine Native Plant Group. Nomenclature followed various sources as cited above. However, priority was given to the Flora Malesiana (van Steenis 1950-ongoing), whenever possible. The legumes were assigned to the traditional family of Leguminosae. Scientific names were not derived from the translation of local names in any case.

Data analysis

Species richness, diversity and evenness were determined for each of the 49 plots. Only those plants rooting within the plots were considered in the

analysis. The Shannon-Index (H') was used as a robust and simple diversity measure (Magurran 1988). For the analysis of species dominance patterns, Evenness (E) based on the Shannon-Index was calculated for each of the plots.

To assess the area's value as a refuge to Philippine tree species, the characterization of forest types by Whitford (1911) was used. His classification and characterization is based on the occurrence of typical tree species and tree species combinations. He often used vernacular names or typical families or genera as e.g. 'Apitong' for *Dipterocarpus* spp. to characterise his forest types. For many of these vernacular names a scientific species could not be assigned with certainty, and therefore, were not used for comparisons. Whitford (1911) pointed out that the description of his forest types was based on a still fragmentary knowledge of Philippine forests. Most of his 'typical' tree species – with few exceptions such as mangroves – occur in the other forest types as well. For example, many species of the dipterocarp forest types occur at wet localities in the Molave forest (limestone forest). On the other hand, the typical Molave forest species also exist in the dipterocarp forest types, especially on dry sites. Of such reasons, Whitford's (1911) forest types are primarily related to the major habitat conditions in the Philippines and do not represent real plant associations. The comparison of the species recorded in this study with Whitford's (1911) forest types merely demonstrates the diverse habitat conditions in the present study area. Unfortunately, not much work has been conducted so far to improve Whitford's system, and information on species composition of the undergrowth vegetation, which might be especially valuable to characterise habitat conditions (Schulze and Whitacre 1999), is still missing.

Results

From the 49 plots, a total of 685 taxa was recorded. Of these, 58.3% were identified to species level, 86.2% to genus level, and 96.7% to family level. The remaining 3.3% of the taxa could only be assigned to higher taxonomic levels. All taxa identified to species level are listed in the Appendix. Species inventory was clearly dominated by angiosperms, accounting for 92.1% of all species. The pteridophytes represented 7.5% of the species. Only three species of gymnosperms (*Podocarpus rumphii*, *Gnetum gnemon*, *G. latifolium*) were found (Table 1).

More than half (52%) of all species identified are endemic to the Philippines, including one endemic genus (*Greeniopsis*, Rubiaceae). The most common families were the Rubiaceae (35 species) and the Euphorbiaceae (32 species), followed by the herbaceous family of Araceae and the erect and climbing palms (Arecaceae) with 28 species each. The Meliaceae and Moraceae included 27 species each (Figure 2). The ratio between the number of genera and the number of species ranged between 1:1.5 (Anacardiaceae) and 1:6.7 (Moraceae).

The frequency of taxa was low. Nearly half (48.5%) of all taxa were recorded from only one of the 49 plots, and nearly one third (30.5%) of the taxa were

Table 1. Taxonomic composition of 49 non-contiguous plots (100 m² each) in lowland forest remnants of the study area in the foothills of Leyte Cordillera.

	Spermatophytes (%)		Pteridophytes (%)	Total (%)
	Gymnosperms	Angiosperms		
Families	2 (1.8)	94 (84.7)	15 (13.5)	111 (100)
Genera	2 (0.7)	261 (90.3)	26 (9)	289 (100)
Species	3 (0.4)	631 (92.1)	51 (7.5)	685 (100)

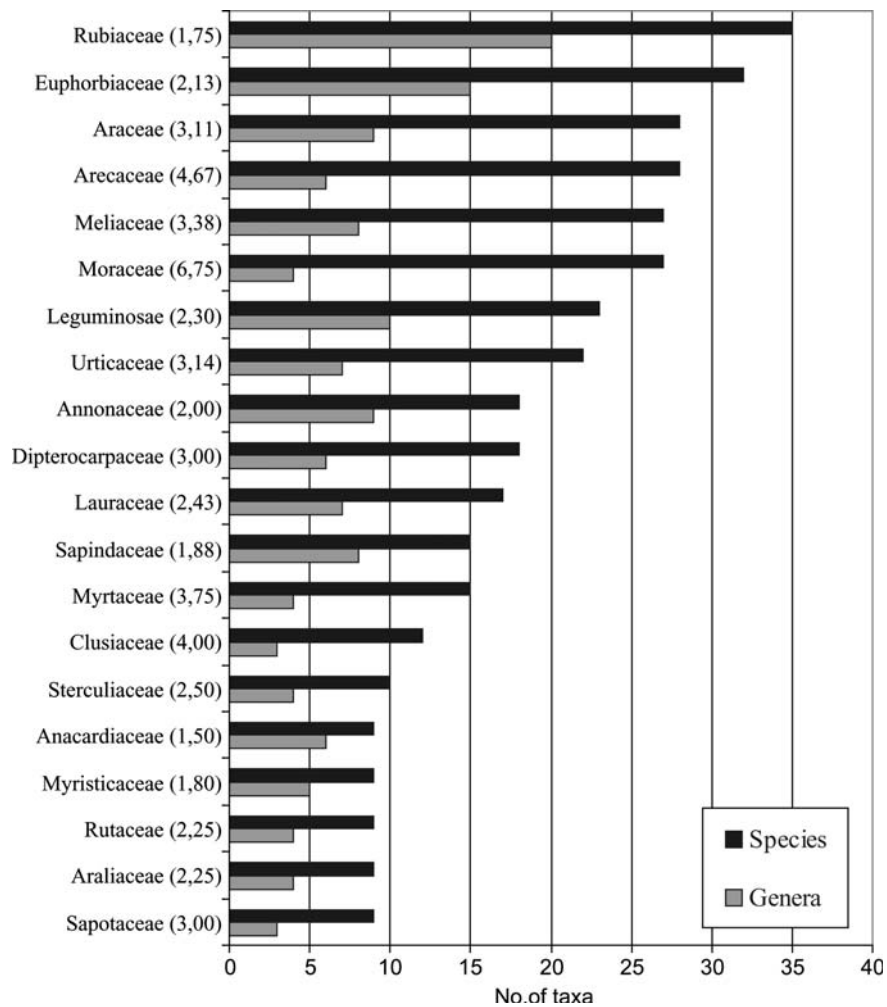


Figure 2. The 20 most common plant families recorded from 49 plots (100 m² each) in the study area. Figures in brackets indicate the ratio between the number of genera and the number of species.

represented by only one single individual. Very few species showed high frequencies as e.g. the two tree species, *Calophyllum blancoi* (present in 32 plots) and *Dacryodes rostrata* (present in 31 plots), which was due to a high rate of juveniles.

The average number of species per plot was 47 and ranged between 17 and 80. Shannon diversity (H') reached values between 2.2 and 3.9, and evenness (E) ranged between 0.64 and 0.98. The species–area curve for all plots shows a steady increase of species numbers with only a weak tendency to level off (Figure 3). The flattening of the curve at its beginning is the result of the river bank vegetation which was comparatively species poor and homogenous. The species–area curve starts to rise again with the addition of the slope plots.

Life form composition is clearly dominated by phanerophytes (65.3% of all taxa), followed by lianas (17.1%) and chamaephytes (16.9%). Geophytes were rare (0.7%) and largely represented by few ground orchids. Hemicryptophytes and therophytes were absent (Figure 4). Epiphytes were not the focus of this study and are therefore not included in the calculation of life form composition. A rough estimate of epiphyte contribution to the area's species inventory is ca. 10%. The most conspicuous epiphytic plant group observed were orchids. Many of the vegetation clusters observed in the tree crowns were composed of the accumulation of orchid bulbs belonging to a single species (e.g. *Grammatophyllum multiflorum*).

The following taxa occurring in the study plots have been classified by Soepadmo (1995) as endangered and economically important lowland forest genera in SE Asia: *Anisoptera*, *Dipterocarpus*, *Parashorea*, *Shorea*, *Vatica* (Dipterocarpaceae), *Artocarpus* (Moraceae), *Mangifera* (Anacardiaceae), and *Calamus* (Arecaceae). Additionally, 41 of the tree species recorded are listed as endangered for the Philippines by IUCN (2000). Of these, 23 are classified as vulnerable, one as endangered, and 17 as critically endangered (see Appendix).

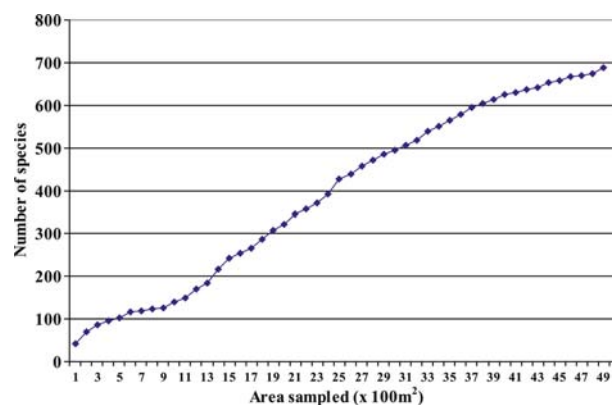


Figure 3. Species–area curve for 49 plots (100 m² each) in the study area.

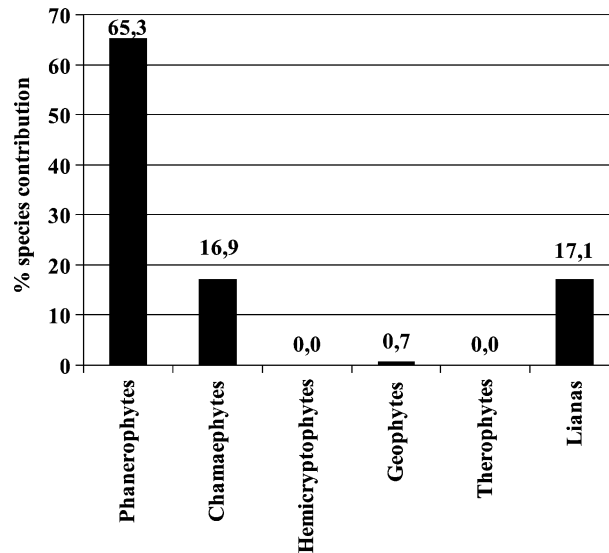


Figure 4. Life form spectrum (after Ellenberg and Müller-Dombois 1967) of species recorded from 49 plots (100 m² each) in the study area.

Discussion

The 685 taxa recorded from the 49 plots account for nearly 8% of the ca. 8900 vascular plant species so far described for the Philippines (Davis et al. 1995). Although the plots were not contiguous and species numbers can therefore be expected to be higher than in contiguous plots (Whitmore 1985) this figure is high, considering the small overall study area (4900 m² in total). Only very few datasets cover tropical lowland forest vegetation comprehensively and are therefore suitable for comparison. The only study using the same plot size was conducted by Whitmore et al. (1985) in the tropical lowland rain forest of Costa Rica, who analysed a plot of 100 m², considering all vascular plants. They recorded a total of 233 species, including 59 (25%) epiphyte species. In the present study, the highest number of species recorded from a single 100-m² plot was 80 and thus much lower than the number found by Whitmore et al. (1985). However, in the present study the vegetation up to 2.5 m tall as well as the lianas were collected from subplots of 25 m², and crown epiphytes were excluded. Despite this, the maximum number of vascular plant species on 100-m² plots in the study area can be expected to be clearly lower than the number of 233 species recorded by Whitmore et al. (1985).

An estimate of the overall vascular plant species richness of Mt. Pangasugan area, including mossy forest as well as the different stages of succession, results in 1500–2000 species. This estimate is based on the very conservative assumption that 50% of the lowland forest species was recorded in this study, and that the mossy forest has a similar species richness as a 1-ha plot studied by

Meijer (1959) in a montane rainforest in Indonesia (333 vascular plant species). The numbers of tree species given by Ingle (2001) (100 species ≥ 5 cm dbh) and Hamann et al. (1999) (92 species ≥ 10 cm dbh) for 0.75- and 1-ha plots, respectively, in Philippine mountain environments show that the overall species richness including all life forms can be expected to be roughly similar to that of Meijer (1959) in Indonesia.

Representation of taxa

The composition of taxa observed in this study is similar to other areas in Southeast Asia. Differences to such sites are related to the proportions of families and result mainly from different inventory approaches. For trees alone, the dominance of the Dipterocarpaceae and the Euphorbiaceae concerning number of species is well documented (Manokaran and Kochummen 1990; Sist and Saridan 1998; Slik et al. 2003; Wilkie et al. 2004). Sist and Saridan (1998) report that the Dipterocarpaceae represent 70% of all trees ≥ 50 cm dbh in a primary forest in East Kalimantan. In our study, the Dipterocarpaceae were the most common family in the canopy layer (12 of 44 species).

Turner (1994) analysed the vascular flora of Singapore and its main habitat types from herbarium collections. The Orchidaceae are the most speciose family in his taxonomic spectrum. This reflects the large number of orchid species in Malesia (6500 species according to Soepadmo 1995). In our study, however, Orchidaceae are poorly represented because we did not include crown epiphytes. Without considering the orchids in both studies, the pteridophytes are the most speciose group, followed by the Rubiaceae and the Euphorbiaceae in both studies. The other predominant families in terms of species richness listed by Turner (1994) are Annonaceae, Moraceae, Arecaceae/Palmae, Myrtaceae, Melastomataceae and Lauraceae. With exception of the Melastomataceae, these are also the most speciose families in our study (Figure 2).

Representation of forest types

The rugged relief of the study area represents a broad spectrum of Philippine habitats. The comparison of the tree species recorded from our study with the typical tree species composition of the forest types described by Whitford (1911) showed a high degree of correspondence. Many tree species typical of the five dipterocarp forest types as well as the Molave type (Figure 5) were present. From the 18 tree species listed by Whitford (1911) as typical for the Lauán-hagághak, 15 were also present in our study area. Originally, this forest type is established on lowland plains on wet soils (Whitford 1911), but was transformed into rice fields in the study area. However, tree species representing this type of forest still occur on the banks of the small creeks at low

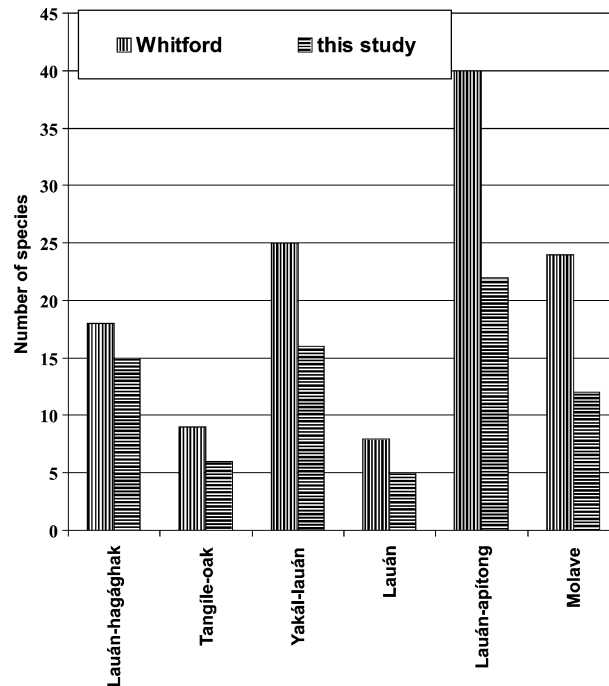


Figure 5. Comparison of the number of characteristic species of the different lowland forest types in the Philippines (after Whitford 1911) with the number of respective species recorded in this study.

elevations. The typical tree species of other forest types were also well represented (Figure 5). The high number of Molave type species (50% of the typical species as mentioned by Whitford 1911) in the study area is remarkable, as this forest represents dry limestone areas (Whitford 1911). This is another indication that the area's vegetation might be strongly influenced by drought periods.

Life form composition

The dominance of trees and phanerophytes is a typical feature of tropical rain forests (Richards 1996). In our study, Phanerophytes account for 65.3% of the species. Richards (1996) provides figures from a rain forest in Guyana, which are based on the Raunkiaer System (Raunkiaer 1934) and cannot be directly compared with our data. We therefore recalculated his data by excluding the lianas from the phanerophytes and excluded the epiphytes in addition. This resulted in a life form composition of 60% phanerophytes, 16% chamaephytes, and 24% lianas. A similar recalculation of figures provided by Cromer and Pryor (1942) for a rain forest in Queensland results in 77.1% phanerophytes, 12.5% chamaephytes, and 10.4% lianas. Figures for a terra firme rain forest in

Brasilia (Cain et al. 1956) are: phanerophytes 74.3%, chamaephytes 0.9%, hemicryptophytes 2.8%, geophytes 0.9%, lianas 12.8%, and epiphytes 8.3%.

Therophytes and hemicryptophytes are usually absent from undisturbed tropical rain forests (Richards 1996). Geophytes are also often absent as in Richards' Guyana study or represented by few species as in the present study (0.7%), where they were mainly made up of ground orchids.

The estimated proportion of epiphyte species of the total number of species in this study (ca. 10%) is clearly lower than the numbers given by Whitmore et al. (1985) (25%) for Costa Rica or by Gentry and Dodson (1987) (35%) for Ecuador.

Conservation value

Kochummen et al. (1992) stated that comparatively small areas might represent high numbers of a regional flora. They found that their 50-ha plot in the Pasoh Forest Reserve (Malaysia) included 25% of all trees and shrubs (≥ 1 cm dbh) of the Malay Peninsula. In our study, an overall sample area of approximately half hectare included ca. 8% of all Philippine vascular plant species. Given the small area considered as well as the fact that neither the successional vegetation nor the mossy forest is included, the representation of Philippine flora in the Mt. Pangasugan area is clearly higher than 8%.

The proportion of 52% endemic taxa recorded in this study is clearly higher than the proportion of 39% stated as an average for the Philippines (Davis et al. 1995). This result agrees with Ashton (1993) who stated that the southeastern part of the Philippines is especially rich in endemic plants. The area's endemism might be even higher than 52%, as a number of taxa could not be identified. For example, only 3 of the 16 rattan species (Arecaceae) recorded, which generally show a high degree of endemism (Dransfield 1990), could be assigned to a scientific name. Two of them were Philippine endemics.

Another aspect referring to the conservation value of the area is the occurrence of 41 tree species in the red list of IUCN (2000). However, from the species recorded from this study, other than trees are not represented in the red list. Despite this, it can be expected that many of the non-tree taxa recorded are threatened by habitat destruction as well. For example, no rattans are listed by IUCN although this plant group is still heavily exploited and shows high rates of endemism. The IUCN red list seems to have a strong focus on well known and economically important tree species. This is supported by the fact that only dipterocarps are classified as critically endangered, although many other tree species are more rare in the study area. This was e.g. true for the valuable tree species *Heritiera sylvatica* (Sterculiaceae) and *Xanthostemon verdugonianus* (Myrtaceae) which were known by local farmers from only one mature tree each in the entire western foothills of Mt. Pangasugan.

Taken together, the Mt. Pangasugan region on Leyte represents a unique refuge for a high number of species, which are characteristic of all Philippine

dipterocarp forest types and the molave type. In view of the large areas of degraded land in the Philippines, the conservation value of the Mt. Pangasugan region is very high and represents an important gene bank of the Philippine forest vegetation.

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Appendix

Species list of the vascular plant species found in 49 plots (100 m² each) in the foothills of the Leyte Cordillera at Mt. Pangasugan, Leyte, Philippines.

The list includes only those species which could be identified to species level. Some species recorded outside the plots are provided in addition.

Numbers in brackets following the species name indicate the first voucher specimen collected of this species.

Life form classification of species is based on observations of mature individuals in the study area, or from species descriptions in literature. Life form definitions follow Ellenberg and Mueller-Dombois (1967) with a minor revision by Richter (1997). MacP, Macrophanerophyte (>20–50 m); MesP, Meso-phanerophyte (>5–20 m); MiP, Microphanerophyte (>2–5 m); NP, Nano-phanerophyte (>1–2 m); NP herb, herbaceous Nanophanerophyte; Ch, Chamaephyte (≤ 1 m); Ch frut, fruticose Chamaephyte; Ch suff, suffruticose Chamaephyte; Ch herb, herbaceous Chamaephyte; G rhiz, rhizome Geophyte; PL, Phanerophytic Liana; r PL, root PL; st PL, winding PL; el PL, tendril PL; d PL, spread climber; E, Epiphyte.

Species classified by IUCN (2000) as endangered are listed along with their status in bold letters. Short definitions of the status are:

CR, critically endangered ('... facing an extremely high risk of extinction in the wild in the immediate future ...'); EN, endangered ('... not critically endangered but facing a very high risk of extinction in the wild in the near future ...'); VU, vulnerable ('... not critically endangered or endangered but facing high risk of extinction in the wild in the medium-term future ...').

For comprehensive definitions and criteria of classification see www.iucnredlist.org/search-basic.html

I.

Spermatophyta	Life form
Aceraceae	
<i>Acer laurinum</i> Hassk. (1221)	MacP
Actinidiaceae	
<i>Saurauia</i> cf. <i>denticulata</i> C.B. Rob. (1078)	MiP
<i>Saurauia samarensis</i> Merr. (235)	MiP
Alangiaceae	
<i>Alangium longiflorum</i> Merr. (1331) VU	MesP
Amaranthaceae	
<i>Deeringia polysperma</i> (Roxb.) Moq. (2214)	Ch herb
Anacardiaceae	
<i>Dracontomelon dao</i> (Blco.) Merr. & Rolfe (660)	MacP
<i>Dracontomelon edule</i> (Blco.) Skeels	Mes
<i>Koordersiodendron pinnatum</i> (Blco) Merr. (162)	MacP
<i>Mangifera altissima</i> Blco. (971) VU	MacP
<i>Rhus taitensis</i> Guill. (818)	MesP
<i>Semecarpus cuneiformis</i> Blco. (538)	MiP
Annonaceae	
<i>Alphonsea arborea</i> (Blco.) Merr. (1009)	MesP
<i>Anaxagorea javanica</i> Bl. (1509)	MiP
<i>Artabotrys</i> cf. <i>rolfei</i> Vid. (2159)	el PL
<i>Cananga odorata</i> (Lamk.) Hook. f. & Thoms.	MesP
<i>Goniothalamus elmeri</i> Merr. (327)	MiP
<i>Meiogyne virgata</i> (Bl.) Miq.	MesP
<i>Papualthia</i> cf. <i>lanceolata</i> (Vid.) Merr. (206)	MesP
<i>Popowia pisocarpa</i> (Bl.) Endl. (1054)	MesP
Apocynaceae	
<i>Alstonia macrophylla</i> Wall. ex. G. Don (1774)	MesP
<i>Alstonia scholaris</i> (L.) R. Br.	MesP
<i>Kibatalia blancoi</i> (Rolfe) Merr. (467)	MesP
<i>Lepiniopsis ternatensis</i> Val. (2220)	MesP
<i>Tabernaemontana pandacaqui</i> Poir. (140)	MicP
<i>Voacanga globosa</i> (Blco.) Merr. (218)	MiP
Araceae	
<i>Alocasia</i> cf. <i>zebrina</i> Schott ex van Houtte (1677)	Ch herb
<i>Amorphophallus paeoniifolius</i> (Dennst.) Nicolson (1924)	NP herb
<i>Costus speciosus</i> (J. König) Sm	NP herb

Spermatophyta	Life form
<i>Pothos cylindricus</i> Presl (1226)	r PL
<i>Raphidophora korthalsii</i> Schott (881)	d PL
Araliaceae	
<i>Arthropodium ahernianum</i> Merr. (1911)	MesP
<i>Osmoxylon trilobatum</i> (Merr.) Philipson (220)	NP
<i>Polyscias nodosa</i> (Bl.) Seem.	MesP
Arecaceae	
<i>Calamus</i> cf. <i>merrillii</i> Becc.	d PL
<i>Caryota</i> cf. <i>cumingii</i> Lodd. ex Mart	MiP
<i>Caryota</i> cf. <i>mitis</i> Lour. (560)	MesP
<i>Daemonorops</i> cf. <i>mollis</i> (Blco.) Merr. (593)	PL
<i>Korthalsia laciniosa</i> Mart. (1120)	d PL
<i>Pinanga maculata</i> Porte	MiP
<i>Caryota rumphiana</i> Mart. var. <i>philippinensis</i> Becc.	MacP
Aristolochiaceae	
<i>Aristolochia philippinensis</i> Warb. (702)	Ch suff
Asclepiadaceae	
<i>Hoya multiflora</i> Bl. (689)	PL
Asteraceae	
<i>Vernonia arborea</i> Buch.-Ham. (826)	MesP
Bignoniaceae	
<i>Oroxylum indicum</i> (L.) Vent.	MesP
<i>Radermachera pinnata</i> (Blco.) Seem. (1129)	MesP
Burseraceae	
<i>Canarium asperum</i> Benth. (265)	MacP
<i>Canarium denticulatum</i> Bl. (428)	MesP
<i>Canarium euryphyllum</i> Perk. (1265)	MacP
<i>Canarium gracile</i> Engl. (611)	MesP
<i>Canarium hirsutum</i> Willd. (1714)	MesP
<i>Dacryodes rostrata</i> (Bl.) H. J. Lam (247)	MacP
Caprifoliaceae	
<i>Sambucus javanica</i> Reinw. ex Bl.	MiP
Casuarinaceae	
<i>Gymnostoma rumphianum</i> (Miq.) L.A.S. Johnson (1915)	MesP
Cecropiaceae	
<i>Poikilospermum erectum</i> (Blco) Merr. (321)	d PL
<i>Poikilospermum suaveolens</i> (Bl.) Merr. (328)	d PL
Celastraceae	
<i>Bhesa paniculata</i> Arn. (812)	MesP
<i>Euonymus cochinchinensis</i> Pierre (495)	MesP
<i>Euonymus javanicus</i> Bl. (232)	MesP
<i>Lophopetalum javanicum</i> (Zoll.) Turcz. (1127)	MacP
Chloranthaceae	
<i>Chloranthus erectus</i> (Buch.-Ham.) Verdc. (1522)	Ch suff
<i>Sarcandra glabra</i> (Thunb.) Nakai (562)	Ch frut

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Chrysobalanaceae	
<i>Maranthes corymbosa</i> Bl. (790)	MacP
Clusiaceae	
<i>Calophyllum blancoi</i> Pl. & Tr. (278)	MesP
<i>Calophyllum soulattri</i> Burm. f. (1250)	MesP
<i>Cratoxylum formosum</i> Benth. & Hook. f. ex Dyer (452)	MesP
Combretaceae	
<i>Terminalia microcarpa</i> Decne. (672)	MacP
<i>Terminalia nitens</i> Presl (481) VU	MesP
Commelinaceae	
<i>Floscope scandens</i> Lour.	Ch herb
<i>Forrestia hispida</i> Less. & A. Rich. (423)	Ch herb
<i>Pollia sorzogoniensis</i> (E. Meyer) Steud.	Ch herb
<i>Pollia thyrsiflora</i> (Bl.) Steud.	Ch herb
<i>Rhopalephora</i> cf. <i>vitiensis</i> (Seem.) Fader (2102)	Ch herb
Connaraceae	
<i>Agelaea borneensis</i> (Hook. f.) Merr. (491)	st PL
<i>Connarus culionensis</i> Merr. (686)	PL
<i>Connarus semidecandrus</i> Jack (623)	PL
<i>Ellipanthus tomentosus</i> Kurz (396)	MesP
Crypteroniaceae	
<i>Crypteronia cumingii</i> (Planch.) Planch. ex Endl. (1693)	MesP
Cunoniaceae	
<i>Weinmannia</i> cf. <i>hutchinsonii</i> Merr. (130)	MesP
Datisceae	
<i>Octomeles sumatrana</i> Miq.	MacP
Dilleniaceae	
<i>Dillenia megalantha</i> Merr. (2007) VU	MesP
<i>Dillenia philippinensis</i> Rolfe VU	MesP
<i>Tetracera fagifolia</i> Bl. (674)	st PL
Dioscoreaceae	
<i>Dioscorea hispida</i> Dennst.	PL
Dipterocarpaceae	
<i>Anisoptera thurifera</i> Foxw. ssp. <i>thurifera</i> (353)	MacP
<i>Dipterocarpus gracilis</i> Bl. (486) CR	MacP
<i>Dipterocarpus validus</i> Bl. CR	MacP
<i>Hopea acuminata</i> Merr. (292) CR	MacP
<i>Hopea malibato</i> Foxw. ex Elm. (20) CR	MacP
<i>Hopea philippinensis</i> Dyer (925) CR	MacP
<i>Hopea plagata</i> (Bleo.) Vid. (305) CR	MacP
<i>Parashorea malaanonan</i> (Bleo.) Merr. (267) CR	MacP
<i>Shorea almon</i> Foxw. (430) CR	MacP
<i>Shorea assamica</i> Dyer forma <i>philippinensis</i> (Brandis) Sym. (269) CR	MacP
<i>Shorea astylosa</i> Foxw. (1796) CR	MacP
<i>Shorea</i> cf. <i>hopeifolia</i> (Heim) Sym. (2110) CR	MakP
<i>Shorea contorta</i> Vid. (1001) CR	MacP

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<i>Shorea falciferoides</i> Foxw. ssp. <i>falciferoides</i> (290) CR	MacP
<i>Shorea guiso</i> (Blco) Bl. (384) CR	MacP
<i>Shorea palosapis</i> (Blco) Merr. (263) CR	MacP
<i>Shorea polysperma</i> (Blco) Merr. (297) CR	MacP
<i>Vatica mangachapui</i> Blco. (528) EN	MacP
Ebenaceae	
<i>Diospyros blancoi</i> A. DC. (163) VU	MacP
<i>Diospyros</i> cf. <i>nitida</i> Merr. (1901)	MiP
<i>Diospyros curranii</i> Merr. (1631)	MesP
<i>Diospyros multibracteata</i> Merr. (598)	MiP
<i>Diospyros pilosantha</i> Blco.	MesP
<i>Diospyros pyrrocarpa</i> Miq. (385)	MesP
Elaeagnaceae	
<i>Elaeagnus triflora</i> Roxb. var. <i>triflora</i> (412)	dPL frut
Elaeocarpaceae	
<i>Elaeocarpus cumingii</i> Turcz. (1123)	MesP
Euphorbiaceae	
<i>Acalypha amentacea</i> Roxb. (254)	NP
<i>Antidesma digitaliforme</i> Tul. (371)	NP
<i>Antidesma nitidum</i> Tul. (268)	MiP
<i>Antidesma tomentosum</i> Bl. (919)	MicP
<i>Aporosa benthamiana</i> Hook. f. (573)	MiP
<i>Baccaurea tetrandra</i> (Baill.) Müll. Arg. (360)	MesP
<i>Bridelia glauca</i> Bl. (233)	MesP
<i>Claoxylon brachyandrum</i> Pax & K. Hoffm. (379)	MesP
<i>Cleistanthus</i> cf. <i>glaber</i> Airy Shaw (628)	MesP
<i>Cleistanthus sumatranus</i> (Miq.) Müll. Arg. (396)	MesP
<i>Codiaeum luzonicum</i> Merr.	MiP
<i>Croton cascarilloides</i> Raeusch. (205)	NP
<i>Drypetes</i> cf. <i>megacarpa</i> (Bl.) Pax & Hoffm. (374)	MiP
<i>Drypetes longifolia</i> (Merr.) Pax et Hoffm. (372)	MiP
<i>Glochidion rubrum</i> Bl. (715)	MiP
<i>Macaranga caudatifolia</i> Elm. (735) VU	MiP
<i>Macaranga grandifolia</i> (Blcol.) Merr. VU	MesP
<i>Macaranga hispida</i> (Bl.) Muell.-Arg.	MiP
<i>Macaranga tanarius</i> (L.) Muell.-Arg.	MesP
<i>Mallotus</i> cf. <i>paniculatus</i> (Lam.) Muell.-Arg. (330)	MesP
<i>Mallotus floribundus</i> (Bl.) Muell.-Arg. (228)	MesP
<i>Mallotus lackeyi</i> Elm. (1800)	MesP
<i>Mallotus philippensis</i> (Lam.) Muell.-Arg. (302)	MesP
<i>Neotrewia cumingii</i> (Muell.-Arg.) Pax & Hoffm. (343)	MesP
<i>Omalanthus populneus</i> (Geisel.) Pax	MiP
<i>Phyllanthus leytenis</i> Elm. (250)	Ch frut
<i>Suregada glomerulata</i> (Hassk.) Jones (287)	NP
Fagaceae	
<i>Lithocarpus buddii</i> (Merr.) A. Camus (15)	MacP
<i>Lithocarpus caudatifolia</i> (Merr.) Rehd. (555)	MesP
<i>Lithocarpus coopertus</i> (Blco) Rehd. (387)	MesP

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Flacourtiaceae	
<i>Casearia</i> cf. <i>mindanaensis</i> Merr. (1675)	P
<i>Casearia grewiaefolia</i> Vent. var. <i>gelonioides</i> (Bl.) Sleum. (794)	MesP
<i>Flacourtia</i> cf. <i>indica</i> (Burm. f.) Merr. (378)	MesP
<i>Osmelia philippina</i> (Turcz.) Benth. (352)	MesP
Flagellariaceae	
<i>Flagellaria indica</i> L. (1128)	el PL
Gesneriaceae	
<i>Cyrtandra angularis</i> Elm. (2212)	Ch herb
<i>Cyrtandra glaucescens</i> Kranzl. (960)	Ch herb
<i>Monophyllaea merrilliana</i> Kranzl. (2027)	Ch herb
<i>Rhynchoglossum obliquum</i> Bl.	Ch herb
Gnetaceae	
<i>Gnetum gnemon</i> L. var. <i>gnemon</i> (375)	MesP
<i>Gnetum latifolium</i> Bl.	PL
Hamamelidaceae	
<i>Sycopsis dunnii</i> Hemsl. (739)	MesP
Hernandiaceae	
<i>Illigera megaptera</i> Merr. (721)	PL
Icacinaceae	
<i>Gomphandra cumingiana</i> (Miers) F.-Vill. (1118)	MesP
<i>Gonocaryum calleryanum</i> (Baill.) Becc. (700)	MesP
<i>Miquelia celebica</i> Bl.	PL
<i>Phytocrine macrophylla</i> (Bl.) Bl. var. <i>macrophylla</i>	PL
<i>Platea excelsa</i> Bl. var. <i>borneensis</i> (Heine) Sleum. (1217)	MesP
Ixonanthaceae	
<i>Ixonanthes petiolaris</i> Bl.	MesP
Juglandaceae	
<i>Engelhardtia serrata</i> Bl. (411)	MacP
Lamiaceae	
<i>Gomphostemma javanicum</i> (Bl.) Bth. (285)	Ch herb
Lauraceae	
<i>Actinodaphne apoensis</i> Merr. (1083)	MesP
<i>Actinodaphne bicolor</i> (Elm.) Merr.	MesP
<i>Actinodaphne</i> cf. <i>multiflora</i> Benth. (833)	MesP
<i>Caryodaphnopsis tonkinensis</i> (Lec.) Shaw (441)	MesP
<i>Cinnamomum mercadoi</i> Vid. (468) VU	MacP
<i>Endiandra coriacea</i> Merr. (1883)	MesP
<i>Litsea garciae</i> Vid. (478)	MesP
<i>Litsea leytenis</i> Merr. (805) VU	MesP
<i>Neolitsea</i> cf. <i>vidallii</i> Merr. (272) VU	MiP

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Leeaceae	
<i>Leea aculeata</i> Bl. ex Spreng.	Mip
<i>Leea guineensis</i> G. Don (255)	MiP
<i>Leea quadrifida</i> Merr. (546)	MiP
Leguminosae	
<i>Azelia rhomboidea</i> (Blco.) Vid. VU	MesP
<i>Albizia procera</i> (Roxb.) Benth.	MacP
<i>Albizia saponaria</i> (Lour.) Bl. ex Miq.	MesP
<i>Archidendron clypearia</i> var. <i>casai</i> (Blco.) I.C. Nielsen (1082)	MesP
<i>Archidendron pauciflorum</i> (Benth.) Nielsen (1852)	MiP
<i>Archidendron scutiferum</i> (Blco.) I.C. Nielsen (323)	MesP
<i>Bauhinia integrifolia</i> Roxb. ssp. <i>cumingiana</i> (Benth.) K. & S.S. Larsen (364)	PL
<i>Dalbergia</i> cf. <i>mimosella</i> (Blco) Prain (1435)	MesP
<i>Desmodium laxum</i> DC. (820)	Ch herb
<i>Entada phaseoloides</i> (L.) Merr.	el PL
<i>Erythrina subumbrans</i> (Hassk.) Merr.	MacP
<i>Euchresta horsfieldii</i> (Lesch.) Benn. (847)	Ch herb
<i>Kingiodendron alternifolium</i> (Elm.) Merr. & Rolfe (357)	MacP
<i>Ormosia calavensis</i> Azaola	MesP
<i>Pterocarpus indicus</i> Willd. VU	MesP
<i>Wallaceodendron celebicum</i> Koord. (395)	MacP
Liliaceae	
<i>Dianella ensifolia</i> (L.) DC.	Ch herb
Loganiaceae	
<i>Fagraea auriculata</i> Jack ssp. <i>auriculata</i> (851)	st PL
<i>Fagraea racemosa</i> Jack ex Wall.	MiP
<i>Strychnos luzoniensis</i> Elm. (748)	el PL
Magnoliaceae	
<i>Magnolia liliifera</i> (L.) Baill. var. <i>angatensis</i> (719)	MesP
Marantaceae	
<i>Donax cannaeformis</i> (Forst. f.) K. Schum. (1006)	MiP
Marattiaceae	
<i>Angiopteris evecta</i> (Forst.) Hoffm. (1188)	NP herb
<i>Marattia pellucida</i> Presl (1444)	Ch herb
Melastomataceae	
<i>Memecylon paniculatum</i> Jack (311)	MiP
Meliaceae	
<i>Aglaia argentea</i> Bl. (642)	MesP
<i>Aglaia costata</i> Merr. (275) VU	MesP
<i>Aglaia elliptica</i> Bl. (1295)	MesP
<i>Aglaia luzoniensis</i> (Vid.) Merr. & Rolfe (511)	MiP
<i>Aphanamixis polystachia</i> (Wall.) R.N. Parker (941)	MesP
<i>Chisocheton ceramicus</i> (Miq.) C. DC. (1184)	MesP
<i>Chisocheton cumingianus</i> (C. DC.) Harms (211)	MesP
<i>Chisocheton pentandrus</i> (Blco.) Merr. (753)	MesP

Spermatophyta	Life form
<i>Dysoxylum arborescens</i> (Bl.) Miq. (664)	MesP
<i>Dysoxylum cumingianum</i> C. DC. (316)	MesP
<i>Reinwardtiodendron humile</i> (Hassk.) Mabb. (965)	MesP
<i>Toona calantas</i> Merr. & Rolfe (918)	MacP
<i>Vavaea amicorum</i> Benth. (273)	NP
<i>Walsura</i> cf. <i>pinnata</i> Hassk. (2082)	MesP
Menispermaceae	
<i>Arcangelisia flava</i> (L.) Merr. (1798)	el PL
Monimiaceae	
<i>Matthaea pubescens</i> Merr. (139)	MiP
Moraceae	
<i>Ficus aurita</i> Bl. (210)	MiP
<i>Artocarpus blancoi</i> (Elm.) Merr. (1701) VU	MacP
<i>Artocarpus elastica</i> Reinw. ex Bl. (697)	MacP
<i>Ficus baletae</i> Merr. (v)	MacP
<i>Ficus benjamina</i> L. (1075)	MacP
<i>Ficus cumingii</i> Miq. var. <i>angustissima</i> (Merr.) Corner (778)	MesP
<i>Ficus fistulosa</i> Reinw. ex Bl. (1307)	MiP
<i>Ficus heteropoda</i> Miq. (425)	MiP
<i>Ficus odorata</i> (Blco.) Merr.	MesP
<i>Ficus pedunculosa</i> Miq. (1780)	MiP
<i>Ficus pseudopalma</i> Blco.	NP
<i>Ficus punctata</i> Thunb. (406)	r PL
<i>Ficus ribes</i> Reinw. ex Bl. (405)	MiP
<i>Ficus ruficaulis</i> Merr.	MesP
<i>Ficus subulata</i> Bl. (646, 1966)	PL
<i>Ficus ulmifolia</i> Lam. (451) VU	MesP
<i>Maclura cochinchinensis</i> (Lour.) Corner (1417)	d PL
<i>Streblus ilicifolia</i> (Vid.) Corner (1700)	MesP
<i>Streblus macrophyllus</i> Bl. (216, 335, 613)	MesP
Myristicaceae	
<i>Endocomia macrocoma</i> (Miq.) W.J.J. de Wilde subsp. <i>prainii</i> (King) W.J.J. de Wilde (479)	MesP
<i>Gymnacranthera farquhariana</i> (Hook. f. & Th) Warb. var. <i>paniculata</i> (A. DC.) R. Schouten (541)	MesP
<i>Horsfieldia</i> cf. <i>costulata</i> (Miq.) Warb. (2002)	MesP
<i>Knema glomerata</i> (Blco.) Merr. (641)	MesP
<i>Knema stellata</i> Merr. (1481)	MesP
<i>Myristica</i> cf. <i>frugifera</i> W. J. J. de Wilde (743) VU	MesP
<i>Myristica</i> cf. <i>philippensis</i> Lam. VU	MesP
<i>Myristica simiarum</i> A. DC cf ssp. <i>simiarum</i> (417)	MesP
Myrsinaceae	
<i>Ardisia pardalina</i> Mez. (815)	Ch frut
<i>Ardisia squamulosa</i> Presl (204) VU	Ch frut
<i>Maesa denticulata</i> Mez (241)	MiP
Myrtaceae	
<i>Acmena acuminatissima</i> (Bl.) Merr. & Perry (503)	MacP

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<i>Syzygium</i> cf. <i>densinervium</i> (Merr.) Merr. (749)	MesP
<i>Syzygium</i> cf. <i>xanthophyllum</i> (C.B. Rob.) Merr.	MesP
<i>Syzygium cumini</i> (L.) Skeel	MesP
<i>Tristaniopsis decorticata</i> (Merr.) P.G. Wilson & J.T. Waterh. (142) VU	MesP
<i>Tristaniopsis micrantha</i> (Merr.) P.G. Wilson & J.T. Waterh. (301)	MesP
<i>Xanthostemon verdugonianus</i> Naves (2209) VU	MacP
Ochnaceae	
<i>Gomphia serrata</i> (Gaertn.) Kanis	MesP
Olacaceae	
<i>Erythralum scandens</i> Bl. (780)	el PL
<i>Strombosia philippinensis</i> (Baill.) Rolfe (380)	MesP
Oleaceae	
<i>Olea borneensis</i> Boerl. (306)	MesP
Opiliaceae	
<i>Champeria manillana</i> (Bl.) Merr. (100)	MesP
<i>Melientha suavis</i> Pierre ssp. <i>suavis</i> (366)	MesP
Orchidaceae	
<i>Calanthe triplicata</i> (Willem.) Ames	G rhiz
<i>Ceratostylis retisquama</i> Rchb. f.B143	E
<i>Cymbidium aliciae</i> Quis. (880)	E
<i>Eulophia zollingeri</i> (Reichb.f.) J.J.Smith	G rhiz
<i>Grammatophyllum multiflorum</i> var. <i>tigrinum</i> Lindley.	E
<i>Lepidogyne longifolia</i> (Bl.) Bl.	G rhiz
<i>Liparis wenzelii</i> Ames	G rhiz
<i>Phalaenopsis hieroglyphica</i> (Rchb. f.) Sweet	E
<i>Robiquetia</i> cf. <i>compressa</i> Schltr.	E
<i>Trichoglottis latisejala</i> Ames	E
<i>Trichoglottis rosea</i> (Lindl.) Ames (1055)	E
Pandanaceae	
<i>Freycinetia</i> cf. <i>philippinensis</i> Hemsl. (1353)	r PL
<i>Freycinetia cumingiana</i> Gaudich. (388, 1234)	r PL
<i>Freycinetia multiflora</i> Merr. (1130)	r PL
<i>Freycinetia vidalii</i> Hemsl. (1352)	r PL
<i>Freycinetia membranifolia</i> Elm. (955)	r PL
Pentaphragmataceae	
<i>Pentaphragma grandiflorum</i> Kurz (457, 458, 1407)	Ch herb
Piperaceae	
<i>Piper abbreviatum</i> Opiz (638)	st PL
<i>Piper halconense</i> C. CD.	st PL
<i>Piper toppingii</i> C. CD. (654, 1143)	st PL
<i>Piper viminale</i> Opiz (1205)	st PL
Pittosporaceae	
<i>Pittosporum resiniferum</i> Hemsl. (448)	Mesp

Spermatophyta	Life form
Poaceae	
<i>Bambusa spinosa</i> Roxb.	MesP
<i>Dinochloa</i> cf. <i>pubiramea</i> Gamble	PL
<i>Dinochloa</i> cf. <i>scandens</i> (Bl.) O. Ktze.	PL
Podocarpaceae	
<i>Podocarpus rumphii</i> Bl. (1520)	MacP
Polygalaceae	
<i>Polygala venenosa</i> Juss. ex Poir. (284, 2011)	Ch herb
<i>Xanthophyllum vitellinum</i> (Bl.) Dietr. (992)	MesP
Proteaceae	
<i>Helicia graciliflora</i> Merr. (1154)	MiP
<i>Helicia loranthoides</i> Presl. (1079)	MesP
<i>Helicia robusta</i> (Roxb.) R. Br. ex Wall. (588)	MiP
Ranunculaceae	
<i>Clematis javana</i> DC. (159, 1997)	PL
Rhamnaceae	
<i>Ventilago dichotoma</i> (Blco.) Merr. (723)	PL
<i>Ziziphus angustifolius</i> (Miq.) Hatusima ex Steenis (488)	MesP
<i>Ziziphus crebrivenosa</i> C.B. Rob. (492, 661)	d PL
Rhizophoraceae	
<i>Gynotroches axillaris</i> Bl. (1538)	MacP
Rosaceae	
<i>Prunus arborea</i> (Bl.) Kalkm. var. <i>arborea</i> (1624)	MesP
<i>Prunus</i> cf. <i>fragrans</i> (Elm.) Kalkm. (795)	MesP
<i>Prunus grisea</i> (Bl.) Kalkm. var. <i>grisea</i> (71, 490)	MesP
<i>Rubus fraxinifolius</i> Poiret (2017)	d PL suff
Rubiaceae	
<i>Boholia nematostylis</i> Merr. (1919)	Ch herb
<i>Canthium gynochthodes</i> Baill. (563)	MesP
<i>Diodia ocyneifolia</i> (Willd.) Brem. (1424)	PL
<i>Diplospora</i> cf. <i>fasciculiflora</i> Elm. (663)	MiP
<i>Dolicholobium philippinense</i> Treteuse (260)	MiP
<i>Greeniopsis multiflora</i> (Elm.) Merr. (279)	MesP
<i>Hedyotis baruensis</i> (Miq.) Val. ex Merr. (329)	Ch herb
<i>Hypobathrum purpureum</i> (Elm.) Merr. (1507)	MesP
<i>Ixora bartlingii</i> Elm. (1060)	Mip
<i>Ixora</i> cf. <i>cumingiana</i> Vidal (509)	MiP
<i>Ixora</i> cf. <i>macrophylla</i> Bartl. (207)	MiP
<i>Ixora longistipula</i> Merr. (1122)	MiP
<i>Ixora macrophylla</i> Barth.	MiP
<i>Ixora salicifolia</i> (Bl.) DC. (288)	NP
<i>Lasianthus</i> cf. <i>obliquinerva</i> Merr. (701)	MiP
<i>Morinda bracteata</i> Roxb. (326)	MiP
<i>Mussaenda philippica</i> A. Rich.	MiP
<i>Mussaenda vidallii</i> Elm. (129)	MiP
<i>Mycetia javanica</i> (Bl.) Korth. (258)	Ch suff
<i>Nauclea subdita</i> (Korth.) Stend. (1958)	MiP

Spermatophyta	Life form
<i>Neonauclea formicaria</i> (Elm.) Merr. (793)	MiP
<i>Neonauclea lanceolata</i> (Bl.) Merr. subsp. <i>gracilis</i> (Vidal) Ridsdale (402)	MesP
<i>Praravinia</i> cf. <i>mindanensis</i> (Elm.) Brem. (289)	NP
<i>Psychotria</i> cf. <i>ixoroides</i> Bartl. ex DC. (515)	st PL
<i>Psychotria membranifolia</i> Bartl. ex DC. (257)	NP
<i>Tarenna cumingiana</i> (Vid.) Elm. (464)	MesP
<i>Tarrenoidea wallichii</i> (Hook. f.) D.D.Tirvengadam & C. Sastre (307)	MesP
<i>Timonius arboreus</i> Merr. (248)	MiP
<i>Uncaria</i> cf. <i>perrottetii</i> (A. Rich.) Merr. (325)	el PL
<i>Uncaria lanosa</i> Wall. f. <i>philippinensis</i> (Elm.) Ridsd. (900)	el PL
<i>Uncaria longiflora</i> (Poir.) Merr. (1300)	el PL
<i>Wendlandia luzoniensis</i> DC. (444)	MesP
<i>Xanthophytum fruticosum</i> Reinw. ex Bl. (1005)	NP
Rutaceae	
<i>Clausena anisum-olens</i> (Blco.) Merr. (605)	NP
<i>Lunasia amara</i> Blco. (158)	NP
<i>Micromelum compressum</i> (Blco.) Merr. (771)	NP
<i>Severinia disticha</i> (Blco) Merr. (398)	NP
Sapindaceae	
<i>Allophyllus cobbe</i> (L.) Raeuschel (823)	MesP
<i>Cubilia cubili</i> (Blco.) Adelh. (586)	MacP
<i>Dictyoneura acuminata</i> Bl. ssp. <i>acuminata</i> (246)	MesP
<i>Dimocarpus fumatus</i> (Bl.) Leenhouts ssp. <i>philippinensis</i> Leenhouts (72)	MesP
<i>Euphorianthus obtusatus</i> Radlk. ex Koord. (1641)	MesP
<i>Ganophyllum falcatum</i> Bl. (1212)	MesP
<i>Guioa</i> cf. <i>diplopetala</i> (Hassk.) Radlk. (1104)	MesP
<i>Harpullia cupanioides</i> Roxb. (212)	MesP
<i>Lepisanthes fruticosa</i> (Roxb.) Leenh. (933)	MesP
<i>Nephelium</i> cf. <i>ramboutan-ake</i> (Labill.) Leenh. (442)	MesP
<i>Paranephelium</i> cf. <i>xestophyllum</i> Miq. (727)	MesP
<i>Pometia pinnata</i> Forst. (578, 1546)	MesP
Sapotaceae	
<i>Palaquium philippense</i> (Perr.) C. B. Rob. (443) VU	MacP
<i>Planchonella mindanaensis</i> Clemens (1126)	MacP
<i>Pouteria firma</i> (Miq.) Baehni (1237)	MacP
Saxifragaceae	
<i>Dichroa philippinensis</i> Schltr.	Ch frut
<i>Polyosma integrifolia</i> Bl. (1219)	NP
<i>Dichroa fibrifuga</i> (807)	
Simaroubaceae	
<i>Picrasma javanica</i> Bl. (218)	MesP
Solanaceae	
<i>Solanum anisophyllum</i> Elm. (225)	Ch herb
<i>Solanum ferox</i> L. (282)	Ch herb
Sonneratiaceae	
<i>Duabanga moluccana</i> Bl.	MacP

Spermatophyta	Life form
Staphyleaceae	
<i>Turpinia borneensis</i> (Merr. & Perry) B.L. Linden (1802)	MesP
Sterculiaceae	
<i>Heritiera sylvatica</i> Vidal (1768)	MesP
<i>Pterocymbium tinctorium</i> (Blco.) Merr. (345)	MacP
<i>Pterospermum diversifolium</i> Bl. (270)	MesP
<i>Pterospermum elongatum</i> Korth. (434)	MesP
<i>Pterospermum obliquum</i> Blco. (120)	MesP
<i>Sterculia multistipularis</i> Elm. (251)	MiP
<i>Sterculia oblongata</i> R. Br. (678)	MesP
<i>Sterculia philippinensis</i> Merr. (1898)	MesP
<i>Sterculia stipulata</i> Korth. var. <i>jagorii</i> (Warb.) Tantra	MesP
Symplocaceae	
<i>Symplocos cochinchinensis</i> (Lour.) Moore var. <i>cochinchinensis</i> (1954)	MicP
Taccaceae	
<i>Tacca palmata</i> Bl. (303)	G rhiz
Theaceae	
<i>Eurya acuminata</i> DC. (1314)	NP
<i>Ternstroemia philippinensis</i> Merr. var. <i>philippinensis</i> (1491)	MesP
Thymelaeaceae	
<i>Aquilaria cumingiana</i> (Decn) Ridl. (300)	NP
<i>Phaleria perrottetiana</i> (Dcne) F.-Vill. (160)	Ch suff
Tiliaceae	
<i>Colona serratifolia</i> Cav. (626)	MesP
<i>Diplodiscus paniculatus</i> Turcz. (271) VU	MesP
Ulmaceae	
<i>Celtis</i> cf. <i>philippinensis</i> Blanco	MesP
<i>Gironniera celtidifolia</i> Gaudich. (238)	MiP
<i>Trema orientalis</i> (L.) Bl. (2202)	MesP
Urticaceae	
<i>Cypholophus moluccanus</i> (Bl.) Miq.	Ch frut
<i>Leucosyke capitellata</i> (Poir.) Wedd. (242)	MiP
<i>Maoutia setosa</i> Wedd.	NP
<i>Villebrunea rubescens</i> (Bl.) Bl. (324)	MesP
<i>Villebrunea trinervis</i> Wedd. (733)	MesP
Verbenaceae	
<i>Clerodendrum villosum</i> Bl.	NP
<i>Teijsmanniodendron pteropodum</i> (Miq.) Bakh. (157)	MesP
<i>Vitex parviflora</i> Juss. (1837)	MacP
<i>Vitex turczaninowii</i> Merr. (705)	MacP
<i>Premna odorata</i> Blco. (633)	MesP

II.

Pteridophyta	Life form
Aspidiaceae	
<i>Didymochlaena</i> cf. <i>truncatula</i> (Sw.) J. Sm. (2056)	Ch herb
Aspleniaceae	
<i>Asplenium nidus</i> L. (1902)	Ch herb
<i>Asplenium tenerum</i> Forst. (2096)	Ch herb
Athyriaceae	
<i>Diplazium asperum</i> (Bl.) Milde (1809)	Ch herb
<i>Diplazium esculentum</i> (Retz.) Sw. (1846)	Ch herb
Cyatheaceae	
<i>Cyathea</i> cf. <i>contaminans</i> (Hook.) Copel.	MesP
Davalliaceae	
<i>Davallia solida</i> (G. Forst.) Sw. (1462)	Ch herb
<i>Davallia trichomanoides</i> Bl. var. <i>lorrainii</i> (Hance) Holttum (222)	Ch herb
Hymenophyllaceae	
<i>Trichomanes javanicum</i> Bl. (1042)	Ch herb
Lindsaeaceae	
<i>Lindsaea lucida</i> Bl. ssp. <i>lucida</i> (533)	Ch herb
<i>Sphenomeris chinensis</i> (L.) Maxon	Ch herb
<i>Tapeinidium pinnatum</i> (Cav.) C.Chr. (1267)	Ch herb
Lomariopsidaceae	
<i>Bolbitis</i> cf. <i>guoyana</i> (Gaudich.) Ching (2016)	Ch herb
<i>Bolbitis guoyana</i> (Gaudich.) Ching	Ch herb
<i>Bolbitis heteroclita</i> (Presl) Ching (1049)	r PL
<i>Lomogramma</i> cf. <i>copelandii</i> Holttum (1851)	r PL
<i>Lomogramma copelandii</i> Holttum	r PL
<i>Teratophyllum arthropteroides</i> (Christ) Holttum (2084)	Ch herb
<i>Teratophyllum</i> cf. <i>articulatum</i> (J. Sm. ex Fée) Mett. (516)	Ch herb
Osmundaceae	
<i>Osmunda banksiaefolia</i> (Pr.) Kuhn (1261, 1392)	Ch herb
Polypodiaceae	
<i>Drynaria quercifolia</i> (L.) J. Sm	E
<i>Leptochilus</i> cf. <i>decurrens</i> Bl.	Ch herb
<i>Microsorium</i> cf. <i>longissimum</i> J. Sm. ex Fée (964)	Ch herb
<i>Microsorium membranifolium</i> (R. Br.) Ching	Ch herb
<i>Microsorium punctatum</i> (L.) Copel. (1821)	Ch herb
<i>Microsorium scolopendria</i> (Burm. f.) Copel. (1445)	Ch herb
<i>Pyrrosia</i> cf. <i>lanceolata</i> (L.) Farwell	PL
<i>Microsorium plukenetii</i> (Presl) M.G. Price (1860)	
Pteridaceae	
<i>Pteris</i> cf. <i>pellucida</i> Presl	Ch herb
<i>Pteris ensiformis</i> Burm. f. (1806)	Ch herb
<i>Pteris longipinnula</i> Wall. (334)	Ch herb
Schizaeaceae	
<i>Lygodium auriculatum</i> (Willd.) Alst. et Holtt. (1974)	st PL
<i>Lygodium circinnatum</i> (Burm. f.) Sw. (1603)	st PL

Ptaridophyta	Life form
Selaginellaceae	
<i>Selaginella</i> cf. <i>involvens</i> (Sw.) Spring (856)	Ch frut
<i>Selaginella</i> cf. <i>springiana</i> Alderw. (1526)	Ch frut
<i>Selaginella cupressina</i> (Willd.) Spring (745)	Ch frut
<i>Selaginella engleri</i> Hieron. (1011)	Ch frut
Taenitidaceae	
<i>Taenitis blechnoides</i> (Willd.) Sw. (1091)	Ch herb
Tectaria group	
<i>Ctenitis</i> cf. <i>silvatica</i> Holttum (939)	Ch herb
<i>Cyclopetlis crenata</i> (Fée) C. Chr. (1807)	Ch herb
<i>Pleocnemia</i> cf. <i>presliana</i> Holttum (1849)	Ch herb
<i>Pleocnemia irregularis</i> (Presl) Holttum (1007)	Ch herb
<i>Tectaria crenata</i> Cav. (1301)	Ch herb
Thelypteridaceae	
<i>Cyclosorus sumatranus</i> (v. Ald. v. Ros.) Ching	Ch herb
<i>Pneumatopteris laevis</i> (Mett.) Holttum (1812)	Ch herb
<i>Pronephrium</i> × <i>xiphioides</i> (Christ) Holttum (498)	Ch herb
<i>Pseudophegopteris paludosa</i> (Bl.) Ching (2093)	Ch herb
<i>Pronephrium granulatum</i> (Presl) Holtt. (997)	Ch herb

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